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Plant Research for Antifertility: Opportunities, Challenges, and an Evidence-to-Translation Roadmap for Responsible Contraceptive Innovation

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Abstract

Plant-derived antifertility agents have been investigated for decades as potential contraceptives for both women and men. Interest has intensified as health systems seek additional nonhormonal options, culturally acceptable methods, and low-cost technologies—while recognizing that contraceptive development must meet strict standards of safety, reversibility, effectiveness, and user acceptability. Plant research offers multi-target biochemical mechanisms: modulation of the hypothalamic–pituitary–gonadal axis, disruption of spermatogenesis or sperm function, inhibition of ovulation, alteration of endometrial receptivity, or local spermicidal effects. However, the field also faces persistent constraints: heterogeneous evidence quality, limited standardization of complex extracts, incomplete compound identification, variable reversibility, and safety concerns. Historical experience with gossypol illustrates both potential and risk—high efficacy but toxicity and incomplete recovery in some users (Qian, 1984; Liu *et al.*, 1987; Liu, 1987; Coutinho, 1988). More recent work on triptonide from *Tripterygium wilfordii* suggests a promising nonhormonal male contraceptive mechanism with reversibility in animal models (Chang *et al.*, 2021), and neem has been reviewed as a multipurpose plant with contraceptive potential (Patil *et al.*, 2021). This article provides a framework synthesis (≤ 2024) that maps plant antifertility mechanisms, summarizes representative evidence across male and female approaches, and proposes a staged roadmap for translation. Results are presented as two conceptual figures (reproductive targets and evidence ladder) and three tables addressing (1) target–assay alignment, (2) challenges with mitigation strategies, and (3) opportunity pathways and institutional actions. We conclude that plant antifertility research can contribute to future contraceptive innovation if it shifts from isolated screening toward standardized, mechanism-aligned, reversible, and ethically governed evidence programs with transparent safety evaluation and partnership-ready translational planning.

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1. Introduction

Contraception is central to reproductive autonomy, maternal and child health, and socio-economic development. Despite a broad method mix, there remain unmet needs: side effects, contraindications, discontinuation, and limited options for men. In parallel, misinformation and distrust can reduce adoption. These realities motivate exploration of new contraceptive mechanisms that are safe, reversible, affordable, and acceptable.

Plant-based antifertility research is often framed as a “natural alternative,” but scientific and regulatory requirements remain stringent. A credible antifertility agent must be evaluated not only for efficacy but also for safety, reversibility, and impacts on sexual behavior, endocrine function, and long-term health. The history of gossypol underscores this: clinical studies demonstrated high antifertility efficacy but also adverse effects and concerns about recovery of spermatogenesis (Qian, 1984; Liu *et al.*, 1987; Coutinho, 1988) ^[19,5,12,13].

Recent nonhormonal male contraceptive development efforts include plant-derived or plant-inspired molecules. Triptonide from *Tripterygium wilfordii* showed reversible male contraceptive effects in mice and nonhuman primates (Chang *et al.*, 2021) ^[3], and broader reviews emphasize that successful nonhormonal methods must be highly specific to avoid off-target toxicity (Nickels *et al.*, 2024) ^[15]. For female contraception, botanical approaches include spermicidal agents (e.g., neem) and agents affecting ovulation or implantation—yet human evidence is limited and safety is paramount (Patil *et al.*, 2021) ^[18].

This article synthesizes evidence up to 2024 to clarify opportunities and challenges in plant antifertility research and to propose an evidence-to-translation roadmap. Guiding questions are: (1) Which reproductive targets are most plausible for plant-based antifertility agents? (2) What challenges commonly prevent translation? (3) What strategies can universities and research institutes adopt to build ethical, rigorous, and scalable evidence programs?

2. Literature Review

2.1. Conceptual foundations: reproductive targets and antifertility mechanisms

Antifertility research targets multiple stages of reproduction. For men, key targets include spermatogenesis, sperm maturation, and sperm function (motility, capacitation, acrosome reaction). For women, targets include ovulation, cervical mucus properties, tubal transport, fertilization, and implantation. Mechanisms can be systemic (HPG axis modulation) or local (e.g., vaginal spermicides). The most attractive candidates are those with high target specificity and reversibility, minimizing long-term endocrine disruption.

2.2. Evidence from historical plant-derived male contraception research

Gossypol from cottonseed represents the most prominent plant-derived male contraceptive explored in large-scale testing. Reviews and clinical reports describe high efficacy but also hypokalemia, fatigue, and concerns about incomplete reversibility for a subset of users (Qian, 1984; Liu *et al.*, 1987; Liu, 1987; Coutinho, 1988) ^[19,5,12,13]. This history provides design lessons: (a) contraceptive efficacy alone is insufficient, (b) safety margins must be wide, and (c) recovery of fertility must be demonstrable and predictable.

2.3. Modern nonhormonal plant-derived leads and the specificity problem

A recent landmark report described triptonide from *Tripterygium wilfordii* as a reversible nonhormonal male contraceptive agent in mice and nonhuman primates (Chang *et al.*, 2021) ^[3]. Mechanistic work suggests effects on sperm morphology/function without systemic toxicity in studied models, but translation requires deeper validation and safety assessment. A 2024 review of nonhormonal male

contraceptive development highlights the need for highly specific targets and robust safety evaluation, noting that many compounds fail due to off-target effects or insufficient reversibility (Nickels *et al.*, 2024) ^[15].

2.4. Female plant-based contraception: spermicides, ovulation, and implantation targets

Plant-based female contraception research includes local spermicidal approaches and systemic targets. Neem (*Azadirachta indica*) has been widely studied and reviewed for contraceptive potential, though mechanisms and human evidence remain incomplete (Patil *et al.*, 2021) ^[18]. Botanical spermicides must balance efficacy with mucosal safety and microbiome impacts; certain surfactant spermicides have been associated with mucosal irritation in prior contexts. Therefore, formulation science and safety surveillance are as important as *in vitro* spermicidal activity.

2.5. Methodological standards: chemistry, reproducibility, and reporting

Plant antifertility studies are vulnerable to irreproducibility because extracts vary by chemotype, processing, and contaminants. Reliable translation requires voucher specimens, metabolomic profiling and dereplication, and transparent reporting of identification confidence. Standards from metabolomics and natural products workflows emphasize QC samples, internal standards, and careful annotation and deposition of spectral data (Fiehn, 2002; Sumner *et al.*, 2007; Wolfender *et al.*, 2019; Qin *et al.*, 2022) ^[7,22,29].

2.6. Governance, ethics, and access-and-benefit sharing

Plant contraceptive research may involve biodiversity resources and traditional knowledge. Ethical and legal compliance under the Nagoya Protocol framework requires prior informed consent and mutually agreed terms and can influence research timelines and partnerships (CBD, 2011; Oberthür & Rosendal, 2014) ^[17]. Ethical engagement is not an optional add-on; it shapes legitimacy, long-term access, and equity. Best practice in ethnopharmacology emphasizes transparent community engagement and avoidance of extractive research (Heinrich *et al.*, 2020) ^[9].

3. Method

This article uses a framework synthesis method. We integrate theoretical and empirical literature published up to 2024 to develop an evidence-informed view of opportunities and challenges in plant antifertility research.

Sources were grouped into six clusters: (1) reproductive biology targets and contraception evidence standards; (2) historical plant-derived contraceptive development (gossypol); (3) modern plant-derived or plant-inspired nonhormonal leads (e.g., triptonide); (4) female botanical contraception including spermicides (e.g., neem); (5) metabolomics and natural products workflows for identification and reproducibility; and (6) governance and ethics under ABS frameworks.

Results are presented as design-oriented guidance. Figure 1 maps reproductive targets and intervention points for antifertility research. Figure 2 presents an evidence-to-translation ladder emphasizing safety, reversibility, standardization, and acceptability. Tables summarize target–assay alignment, challenge-mitigation strategies, and opportunity pathways for higher education institutions.

4. Results and Discussion

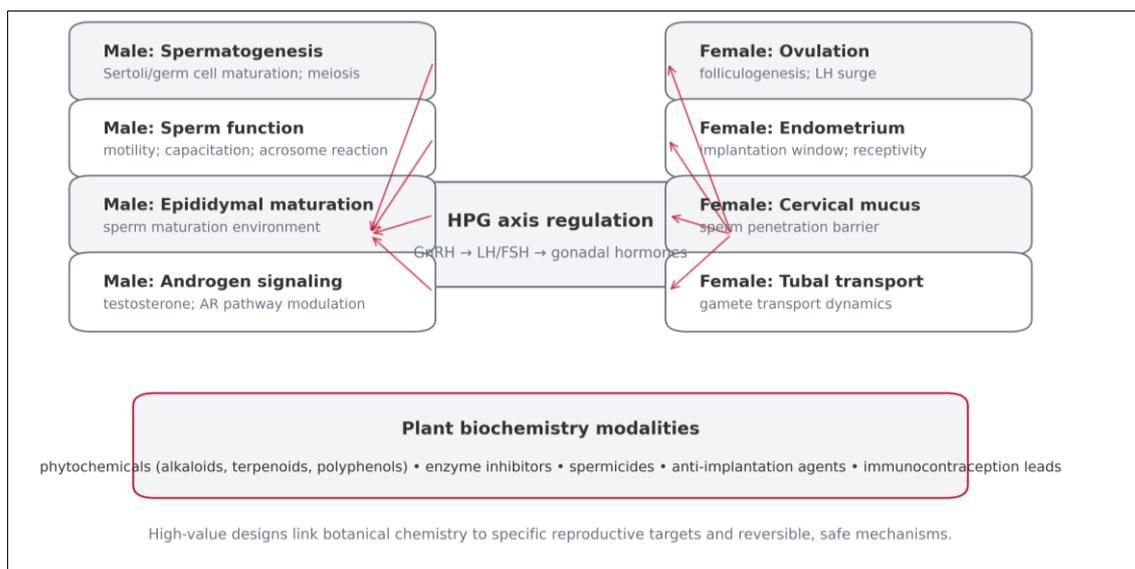


Fig 1: Plant antifertility research: reproductive targets and intervention points



Fig 2: Evidence-to-translation ladder for plant-based antifertility research

Results are organized around mechanism alignment and translation readiness. Figure 1 shows that plant antifertility research can target both systemic endocrine regulation and local reproductive processes, but that the highest translation value comes from specific, reversible mechanisms that avoid long-term endocrine disruption. Figure 2 outlines an evidence ladder for moving from ethnobotanical prioritization to human trials and post-market monitoring.

4.1. Mechanism-aligned research designs.

High-quality antifertility research begins with explicit target definition and assay selection. For male contraception, this includes sperm motility and morphology assays, spermatogenesis markers, and reversibility studies with fertility recovery endpoints. For female approaches, ovulation assays, endometrial receptivity markers, and implantation endpoints may be used, but ethical

considerations are paramount. Local spermicidal formulations require mucosal safety and microbiome assessments.

4.2. Lessons from gossypol:

efficacy is not enough. The gossypol experience demonstrates how a promising plant-derived antifertility agent can fail translation due to toxicity and incomplete recovery of fertility for some users. Clinical trial evidence shows effectiveness but also adverse events and nephrotoxicity concerns (Liu *et al.*, 1987; Liu, 1987; Coutinho, 1988) ^[5,12,13]. This history supports strict go/no-go criteria for new botanical candidates: predictable reversibility, wide safety margins, and well-defined dosing strategies.

4.3. Modern leads and the specificity challenge.

Triptonide illustrates a pathway for plant-derived nonhormonal male contraception with reversible effects in animal models (Chang *et al.*, 2021) [3]. However, a broader 2024 review emphasizes that most nonhormonal leads fail because they interact with essential physiological pathways or have off-target toxicity (Nickels *et al.*, 2024) [15]. Therefore, translation programs should include comprehensive toxicology, pharmacokinetics, and long-term follow-up, not only short-term fertility suppression endpoints.

4.4. Standardization, chemistry, and the reproducibility bottleneck.

A central constraint is inconsistent chemistry. Without vouchers, metadata, and metabolomic fingerprinting, replication across sites and seasons is difficult. Metabolomics and natural products literature emphasize QC practices and transparent identification confidence (Fiehn, 2002; Sumner *et al.*, 2007; Wolfender *et al.*, 2019) [7,22,29]. Molecular networking can accelerate dereplication (Qin *et al.*, 2022), reducing rediscovery and clarifying active components.

4.5. Safety, acceptability, and gender equity.

Contraceptive technologies are only successful if people want to use them. Botanical “naturalness” does not guarantee acceptability; safety concerns and side effects often drive discontinuation. For male methods, reversibility and absence of libido changes are critical. For female local methods, mucosal safety and infection risk are critical. Programs should include user-centered design, acceptability studies, and transparent risk communication.

4.6. Institutional roadmap.

Universities can strengthen plant antifertility research by building (1) core analytics (LC–MS/MS, NMR) and curated spectral libraries, (2) reproductive biology assay platforms and standardized SOPs, (3) governance readiness under ABS frameworks, (4) translational partnerships (clinics, public health agencies, SMEs), and (5) responsible innovation and communication practices. Stage-gates aligned to Figure 2 can prevent overclaims and focus resources on candidates with credible safety and reversibility profiles.

Table 1: Reproductive targets, assays, and evidence needs in plant antifertility research.

Target area	Key endpoints	Recommended assays/models	Critical safeguards
Male spermatogenesis	sperm count; testis histology; hormones	histology; germ cell markers; endocrine panels	reversibility endpoints; endocrine off-target checks
Male sperm function	motility; morphology; acrosome reaction	CASA motility; morphology scoring; capacitation assays	dose–response; cytotoxicity; recovery after washout
Female ovulation	ovulation rate; follicle maturation	cycle tracking in models; hormonal profiling	avoid long-term endocrine disruption; fertility recovery
Endometrium/implantation	receptivity markers; implantation rate	endometrial markers; implantation models	strong ethics; translation caution; avoid overclaiming
Local spermicides	sperm immobilization; mucosal safety	<i>in vitro</i> sperm assays; mucosal irritation models	microbiome and infection risk monitoring
HPG axis modulation	GnRH/LH/FSH; sex steroids	hormonal assays; receptor studies	high off-target risk; prioritize specificity

Table 2: Major challenges and mitigation strategies for plant antifertility research.

Challenge	Why it matters	Mitigation strategy	Evidence anchor (≤2024)
Safety & toxicity	Contraceptives require wide safety margins	Tiered toxicology; long-term follow-up; PK/PD	Nickels <i>et al.</i> (2024) [15]; gossypol experience
Uncertain reversibility	Irreversibility is unacceptable for most users	Washout studies; fertility recovery endpoints	Chang <i>et al.</i> (2021) [3]; gossypol trials
Chemical variability	Batch inconsistency undermines efficacy/safety	Vouchers; metabolomic fingerprints; markers	Fiehn (2002) [7]; Sumner <i>et al.</i> (2007)[22]
Annotation bottleneck	Weak mechanism claims and rediscovery	Dereplication; molecular networking; libraries	Wolfender <i>et al.</i> (2019); Qin <i>et al.</i> (2022) [29]
Governance/ABS	Legal and ethical legitimacy	PIC/MAT templates; benefit-sharing plans	CBD (2011); Oberthür & Rosendal (2014) [17]
Acceptability	Low uptake if users dislike method	User-centered design; acceptability trials	Contraceptive development best practice

Table 3: Opportunity pathways and institutional actions to improve translation (higher education focus).

Opportunity	Institutional actions	Example outputs	Expected benefit
Nonhormonal male methods	Reproductive assay platform; target discovery	Validated sperm-function assays; candidates	Expanded method mix; shared responsibility
Botanical spermicides	Formulation + safety labs; microbiome monitoring	Safe prototypes; irritation data	Localized, user-controlled methods
Standardized botanical products	QA/QC pipelines; marker panels	Standard extracts; fingerprints	Reproducibility and regulatory readiness
Open spectral libraries	Build and share MS/MS and NMR libraries	Local databases; deposited spectra	Faster identification; less rediscovery
Ethical bioprospecting	ABS office; community partnerships	PIC/MAT records; benefit-sharing reports	Long-term access and legitimacy

5. Conclusion

Plant antifertility research presents both opportunities and risks. Opportunities include new nonhormonal contraceptive mechanisms, culturally contextualized options, and low-cost local production. Challenges include chemical variability, incomplete identification, limited standardization, uncertain reversibility, and safety concerns—as illustrated by gossypol's history. Modern leads like triptonide demonstrate renewed potential but require rigorous translational pathways.

This framework synthesis provides a practical toolkit: a target map (Figure 1), an evidence ladder (Figure 2), and tables summarizing assays, challenges, and institutional actions. The key recommendation is a shift from isolated screening to mechanism-aligned, standardized, ethically governed, and user-centered programs with strong toxicology and reversibility evidence. With such programs, plant antifertility research can contribute to responsible contraceptive innovation in Asia and beyond.

6. References

- Anastas PT, Warner JC. Green chemistry: theory and practice. Oxford: Oxford University Press; 1998.
- Convention on Biological Diversity. Nagoya Protocol on Access to Genetic Resources and the Fair and Equitable Sharing of Benefits Arising from Their Utilization. Montreal: Secretariat of the Convention on Biological Diversity; 2011.
- Chang Z, Qin W, Zheng H, *et al.* Triptonide is a reversible non-hormonal male contraceptive agent in mice and non-human primates. *Nat Commun.* 2021;12:1253. doi:10.1038/s41467-021-21517-5
- Chiba T, *et al.* The prevalence of dietary supplements that claim estrogen-like effects. *Nutrients.* 2022;14(21):4501.
- Coutinho EM. Clinical experience with gossypol in non-Chinese men. *Contraception.* 1988;37(3):269-279.
- Facione PA. Critical thinking: a statement of expert consensus (The Delphi Report). Millbrae (CA): California Academic Press; 1990.
- Fiehn O. Metabolomics—the link between genotypes and phenotypes. *Plant Mol Biol.* 2002;48:155-171.
- Goldberg E. A plant root extract, triptonide, is a reversible male contraceptive in mice and monkeys. *Asian J Androl.* 2021;23(6):563-564.
- Heinrich M, Jalil B, Potocnik T, *et al.* Best practice in ethnopharmacology and ethnobotany: a review and recommendations. *J Ethnopharmacol.* 2020;249:112376.
- Huang CW, Lin C, Nguyen MK, *et al.* Biosensor for environmental monitoring and SDGs. *Bioengineered.* 2023;14(1):58-80.
- Julaehaa E, *et al.* Antifertility compound from the seeds of *Carica papaya*. *Procedia Eng.* 2015;148:995-1001.
- Liu GZ. Experiences with gossypol as a male pill. *Am J Obstet Gynecol.* 1987;156(2):485-489.
- Liu GZ, Lyle KC, Cao J. Clinical trial of gossypol as a male contraceptive drug. Part I: efficacy and safety. *Fertil Steril.* 1987;48(3):459-461.
- Murkies AL, Lombard C, Strauss BJ, *et al.* Phytoestrogens. *J Clin Endocrinol Metab.* 1998;83(2):297-303.
- Nickels L, *et al.* Nonhormonal male contraceptive development—strategies and progress. *Pharmacol Rev.* 2024;76(4).
- Nita S, *et al.* Papaya (*Carica papaya* L.) seed extract as male contraception in rats. *Bioscientia Medicina.* 2020;4(2).
- Oberthür S, Rosendal GK. Global governance of genetic resources: access and benefit sharing after the Nagoya Protocol. London: Routledge; 2014.
- Patil SM, *et al.* Azadirachta indica (neem) as a contraceptive: a review. *Phytomedicine.* 2021;85:153540. doi:10.1016/j.phymed.2021.153540
- Qian SZ. Gossypol: a potential antifertility agent for males. *Annu Rev Pharmacol Toxicol.* 1984;24:329-360.
- Qin GF, Zhang X, Zhu F, *et al.* MS/MS-based molecular networking for natural products dereplication. *Molecules.* 2023;28(1):157. doi:10.3390/molecules28010157
- Rickinson M, *et al.* A review of research on outdoor learning. Slough: National Foundation for Educational Research; 2004.
- Sumner LW, Amberg A, Barrett D, *et al.* Proposed minimum reporting standards for chemical analysis. *Metabolomics.* 2007;3:211-221.
- United Nations Population Fund. State of world population report 2022. New York: UNFPA; 2022.
- United Nations. Transforming our world: the 2030 agenda for sustainable development (A/RES/70/1). New York: United Nations; 2015.
- World Health Organization. Quality control methods for herbal materials. Geneva: WHO; 2011.
- World Health Organization. Medical eligibility criteria for contraceptive use. 5th ed. Geneva: WHO; 2015.
- World Health Organization. Family planning: a global handbook for providers (2018 update). Geneva: WHO; 2018.
- Wiryanawan RA, *et al.* Papaya seed extract lowers sperm concentrations, motility and viability in male mice; 2016.
- Wolfender JL, Litaudon M, Touboul D, Queiroz EF. Innovative strategies for natural product dereplication. *Nat Prod Rep.* 2019;36(6):855-868.